

Epidemiologie und mikrobiologische Diagnostik der Sepsis

K. Becker & E. Idelevich

Institut für Medizinische Mikrobiologie, Universitätsklinikum
Münster



Bad Honnef-Symposium

Empfehlungen zur kalkulierten Initialtherapie bakterieller Erkrankungen bei Erwachsenen

Bonn, 21.-22. März 2016

Block C

„Et blieb nix wie et wor“

Sepsis; Definition Version „X“

„Et blieb nix wie et wor“

The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3)

JAMA. 2016;315(8):801-810. doi:10.1001/jama.2016.0287

Sepsis is not a specific illness but rather a syndrome encompassing a **still-uncertain pathobiology**.

- Word count: 9506
- Occurrence of the term:
 - „**pathogen**“ or similar: n = 3 (=0,03%)
 - „**infection**“: n = 65 (=0,68%)



- **BK-Diagnostik zu langsam für primäre Sepsis-Therapie**
- **nur in max. 30-40% der Sepsisfälle Erregernachweis**

Box 2. Key Concepts of Sepsis

- Sepsis is the primary cause of death from infection, especially if not recognized and treated promptly. Its recognition mandates urgent attention.
- Sepsis is a syndrome shaped by pathogen factors and host factors (eg, sex, race and other genetic determinants, age, comorbidities, environment) with characteristics that evolve over time. What differentiates sepsis from infection is an aberrant or dysregulated host response and the presence of organ dysfunction.
- Sepsis-induced organ dysfunction may be occult; therefore, its presence should be considered in any patient presenting with infection. Conversely, unrecognized infection may be the cause of new-onset organ dysfunction. Any unexplained organ dysfunction should thus raise the possibility of underlying infection.
- The clinical and biological phenotype of sepsis can be modified by preexisting acute illness, long-standing comorbidities, medication, and interventions.
- Specific infections may result in local organ dysfunction without generating a dysregulated systemic host response.

Epidemiologie

Estimated population prevalence of sepsis and severe sepsis in Germany (“pre-DRG era”)

Prospective, observational, cross-sectional 1-day point-prevalence study in 2003
(454 ICUs from a representative nationwide sample of 310 hospitals)

(including septic shock)

Characteristic	Sepsis %	95% CI	Severe sepsis %	95% CI
3,877 patients included				
Overall	12.4	10.9–13.8	11.0	9.7–12.2
Stratum				
1 (non-university, ≤ 200 beds)	9.8	7.2–12.5	6.0	3.9–8.1
2 (non-university, 201–400 beds)	11.5	9.1–13.8	9.5	7.3–11.6
3 (non-university, 401–600 beds)	15.5	11.7–19.2	9.0	6.0–12.0
4 (non-university, > 600 beds)	14.5	10.2–18.8	15.3	11.4–19.2
5 (university)	10.5	3.9–17.2	19.3	13.1–25.4
Gender				
Male	11.5	9.5–13.5	11.9	10.1–13.6
Female	13.5	11.5–15.6	9.7	8.3–11.1
Age (years)				
< 50	12.0	9.0–15.0	11.2	8.4–14.0
50–65	12.4	9.7–15.0	12.2	9.6–14.8
> 65	12.6	10.7–14.4	10.4	8.9–11.9

Number of newly diagnosed cases of

- **Sepsis**: approx. 79,000/a = 116/100,000 inhabitants
- **Severe sepsis**: 75,000 /a = 110/100,000 inhabitants

} **approx. 154,000 cases of sepsis/a in Germany**

Hospital mortality of patients with severe sepsis in Germany

Characteristic	Hospital mortality ^a	
	Percentage	95% CI
Overall	55.2	50.2–60.2
Stratum		
1 (non-university, ≤ 200 beds) ^c	48.8	34.6–63.3
2 (non-university, 201–400 beds)	52.0	42.3–61.7
3 (non-university, 401–600 beds)	46.8	33.0–60.8
4 (non-university, > 600 beds)	60.6	50.8–69.7
5 (university)	60.0	50.0–69.3
Gender		
Male ^c	56.5	49.9–62.8
Female	53.8	46.0–61.5
Age (years; tertiles) ^d		
18–59	46.0	37.5–54.7
60–72	60.8	52.2–68.7
≥ 73	58.6	49.9–66.8
Septic shock		
No ^c	47.3	40.3–54.5
Yes	62.4	55.2–69.2

Engel et al.,
Intensive Care Med
(2007) 33:606–618

SEPNET

Fallzahlen, Sterbefälle, Häufigkeits- und Sterblichkeitsraten der Sepsis in Deutschland, 2013 (lt. DRG-Statistik)

Sepsis
inkl. schwere
Sepsis u.
septischer Schock

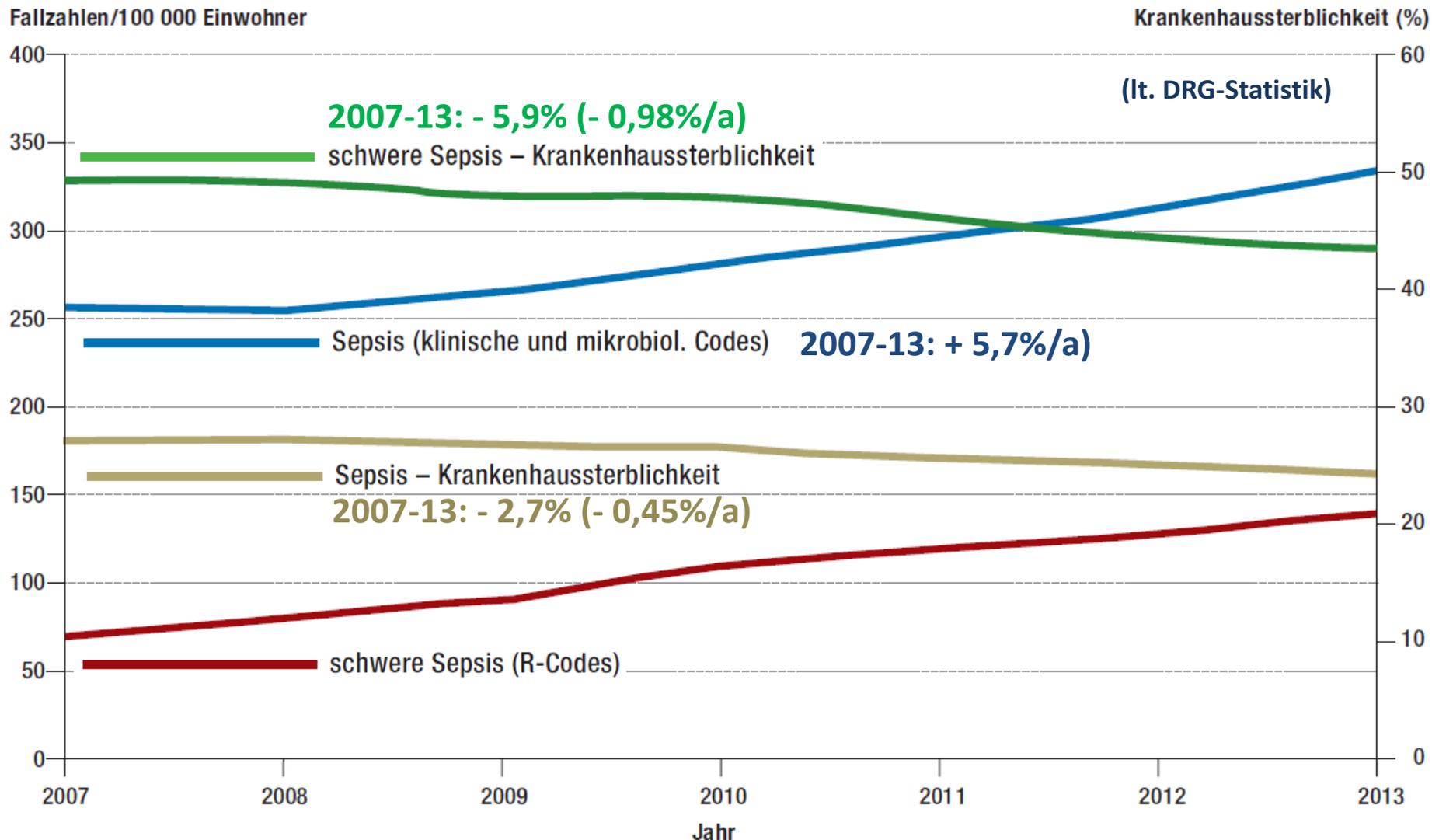
**schwere
Sepsis**
inkl. septischer
Schock
(R65.1!, R57.2)
(% Sepsis, gesamt)

**septischer
Schock**
(R57.2)

(% Sepsis, gesamt)

	Sepsis	schwere Sepsis	septischer Schock
Fälle	279.530	115.421 (41,3%)	33.815 (12,1%)
Todesfälle	67.849	50.349 (74,2%)	19.891 (29,3%)
adjustierte Rate/ 100 000 Einwohner	335	138	40
Krankenhaus- sterblichkeit (%)	24,3	43,6	58,8

Häufigkeitsrate pro 100 000 Einwohner standardisiert auf die Bevölkerungsstruktur 2010 und Krankenhausletalität von Sepsis und schwerer Sepsis (inklusive septischem Schock) in Deutschland, 2007–2013



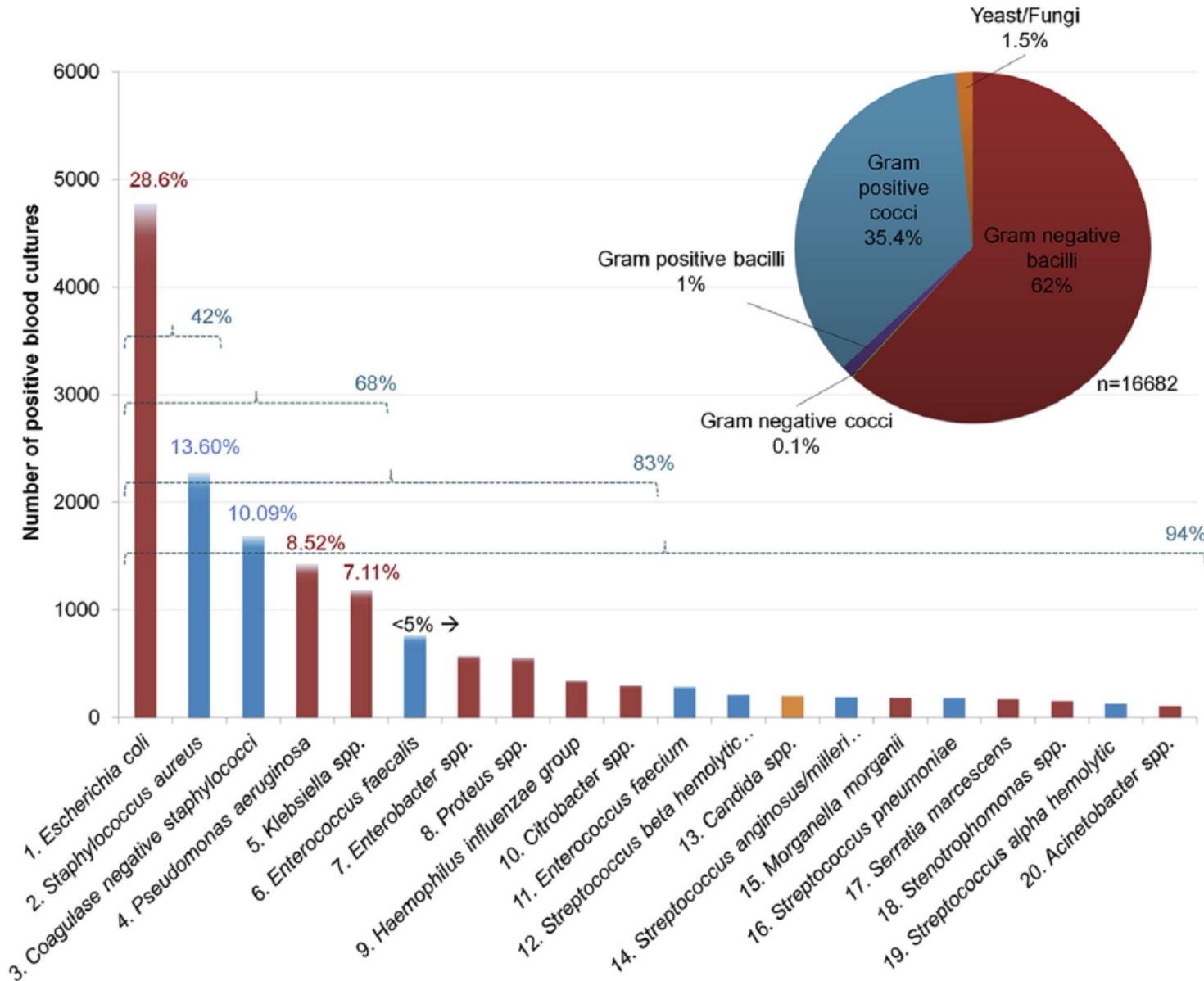
1,21%

↔ Sepsisfälle/Krankenhausfälle ↔

1,54%

Causative pathogens of BSIs

Opota et al., CMI 2015



Top 20 microbes identified from positive blood cultures during 1 year.

Data from our 1000-bed tertiary-care university hospital (Lausanne, Switzerland) during the year 2013 (16 682 identifications).

Das Problem

The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3)

JAMA. 2016;315(8):801-810. doi:10.1001/jama.2016.0287

Identified Challenges and Opportunities

Assessing the Validity of Definitions When There Is No Gold Standard

Sepsis is not a specific illness but rather a syndrome encompassing a still-uncertain pathobiology. At present, it can be identified by a constellation of clinical signs and symptoms in a patient with suspected infection. **Because no gold standard diagnostic test exists,** the task force sought definitions and supporting clinical criteria that were clear and fulfilled multiple domains of usefulness and validity.

110 Jahre Blutkulturdiagnostik



Emanuel Libman
(1872-1946)

- Studie mit >700 Blutkulturen (1906)
- erste Richtlinie zur Blutkulturdiagnostik

JULY, 1906.]

JOHNS HOPKINS HOSPITAL BULLETIN.

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of Batisbon; Marshal Macdonald, who won Wagram, and was made a Marshal on the field of battle, a man who was a gallant and loyal gentleman all his days; Marshal Suchet, who had been a junior officer in Napoleon's company, when the latter was a captain; Marshal Davout, the hero of Auerstadt; General Junot, whose wife Napoleon wished to marry, when she was a young girl—Junot, reckless to gallantry, who resembled Mad Anthony Wayne of our Revolutionary struggle; and General Foy, who was sent to Turkey to fight the Russians

and the English, and was wounded fifteen times in his various arduous campaigns. It seems a pity that Larrey's old friend and patient, Lannes, is not near him. This celebrated Marshal's body lies in the Cemetery of Montmartre.

Those of us that go to Paris and look at the tomb of Baron Larrey can feel with justice that there is buried a soldier, a patriot; a great, learned, and brilliant surgeon; a brave, truthful, and loyal man; a gentleman; and a benefactor of the human race.

ON SOME EXPERIENCES WITH BLOOD-CULTURES IN THE STUDY OF BACTERIAL INFECTIONS.¹

BY E. LIBMAN, M. D.,

Adjunct Visiting Physician and Assistant Pathologist
Mt. Sinai Hospital, New York.

When, a few months ago, you honored me by asking me to present something here, I was much concerned. I felt unequal to the task of addressing you, and I was at a loss as to the choice of my subject. After a few days' thought I decided that the subject which has been announced to you would be an advantageous one in a number of ways.

In the first place, I could combine the paper with a demon-

prefer to speak at this time of our own experiences. The subject of infections by the typhoid bacillus will be omitted and only such pneumococcus infections as occurred independently of pneumonia will be included.

While the paper will not be a connected one,—the breadth of the subject prevents that—I shall take up the following main points:

The organisms found were: streptococci, the pneumococcus, *Micrococcus aureus*, *Micrococcus citreus*, *Micrococcus albus*, *Bacillus pyocyaneus*, paracolon bacillus, *Bacillus coli*, *Bacillus proteus vulgaris* and the gonococcus.

Blood of a child suffering from streptococcal intestinal infection (1), in the clinic of my friend and teacher, Professor Escherich, I have been much interested in systemic bacterial infections, and, during the past eight years I have attempted from time to time to study especially the general infections by the pyogenic cocci. The number of cases which we have studied by means of cultures is between 700 and 750. Not all are of value. Many were made at request in cases which proved to be examples of fever due to tuberculosis, or cases not bacterial in origin. Some were unsuccessful for one reason or another. I shall not attempt to cover the entire material—that would be impossible. Only certain points will be taken up and digressions will be made here and there so as to mention observations that are unusual or particularly interesting. The literature (2) will not be discussed as I

8. Discussion of positive blood-cultures.

9. Diagnostic points.

10. The prognosis in cases in which bacteria have been found in the blood.

1. TERMINOLOGY.

The terminology (3) that I shall use is one based on the work itself and will, I hope, be justified by the observations I shall record. When bacteria attack any part of the body we call the resultant lesion a local infection. There may or may not be evident toxemia; usually there is. If such a local infection is so situated that it is not diagnosed we can call the lesion a cryptic infection. If there is toxemia we have a cryptogenic toxemia.

If bacteria are present in the blood we have a bacteriemia, or systemic infection. Such a bacteriemia may or may not be accompanied by the establishment of secondary foci (metas-

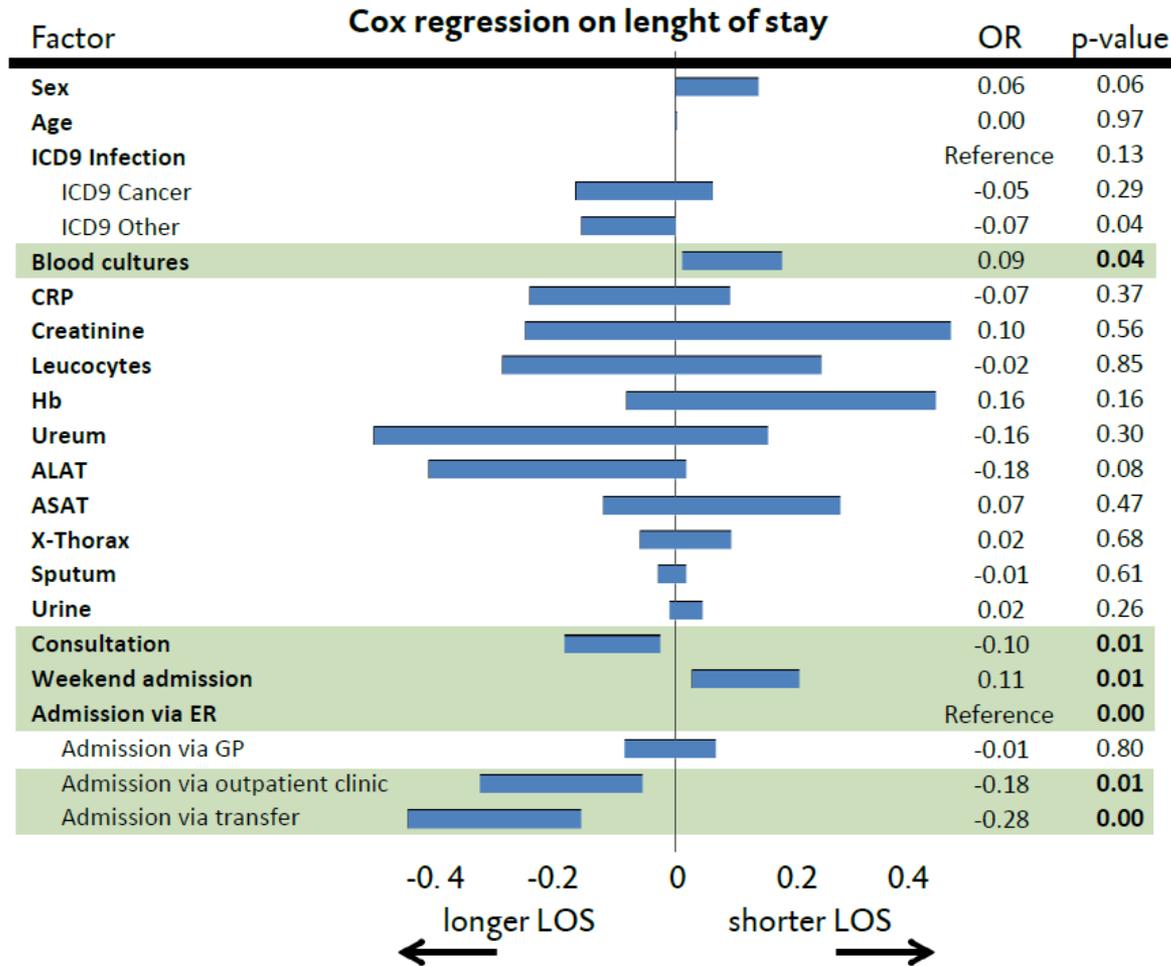
¹Read before the Medical Society of the Johns Hopkins Hospital, February 5, 1906.

dis-
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Effects of doing blood cultures

JH Dik¹, JR Lo Ten Foe¹, B Sinha¹, P Nannan Panday², R Hendrix^{1,3}, MJ Postma⁴ & AW Friedrich¹

¹Department of Medical Microbiology, University Medical Center Groningen, ²Clinical Pharmacy, University Medical Center Groningen, ³Certe, Groningen, ⁴Department of Epidemiology, University of Groningen



Performing blood cultures during admission for patients with a suspected infection was **correlated with:**

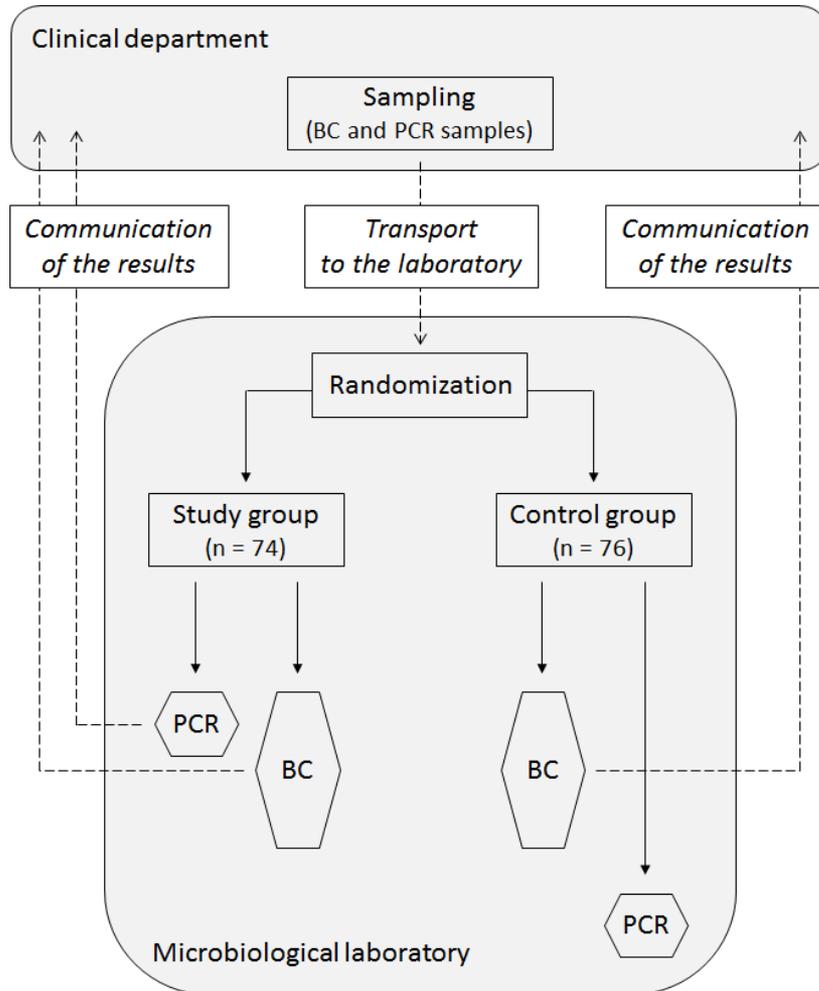
- **Significant shorter stay in the hospital** (16.85 days vs. 13.35 days; $P < 0.001$)
- **Less total costs per patient (€4,788); i.e. saving of €981,441/a for the University Hospital**

Patients had to be started with broad spectrum antibiotics during admission with a minimum of 3 days.

Patients with (n=580) and without (n=205) blood cultures drawn at the day of admission.

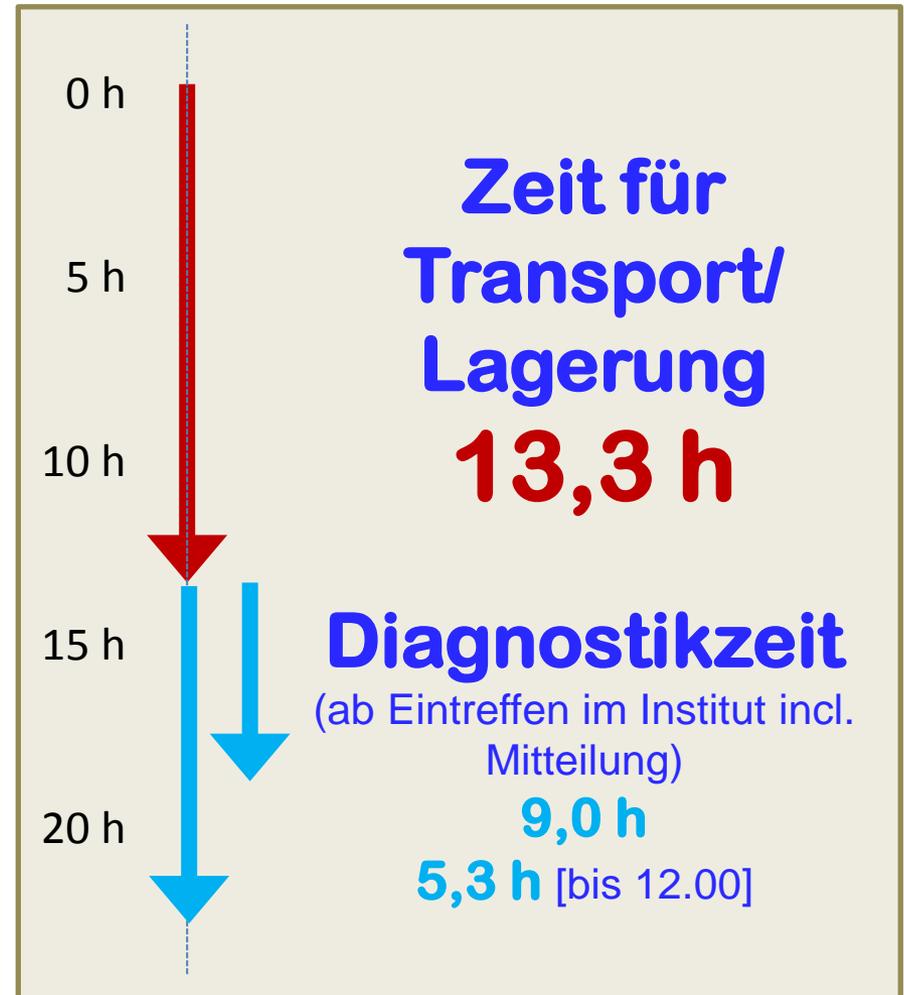
Impact of multiplex PCR on antimicrobial treatment in febrile neutropenia: a randomized controlled study

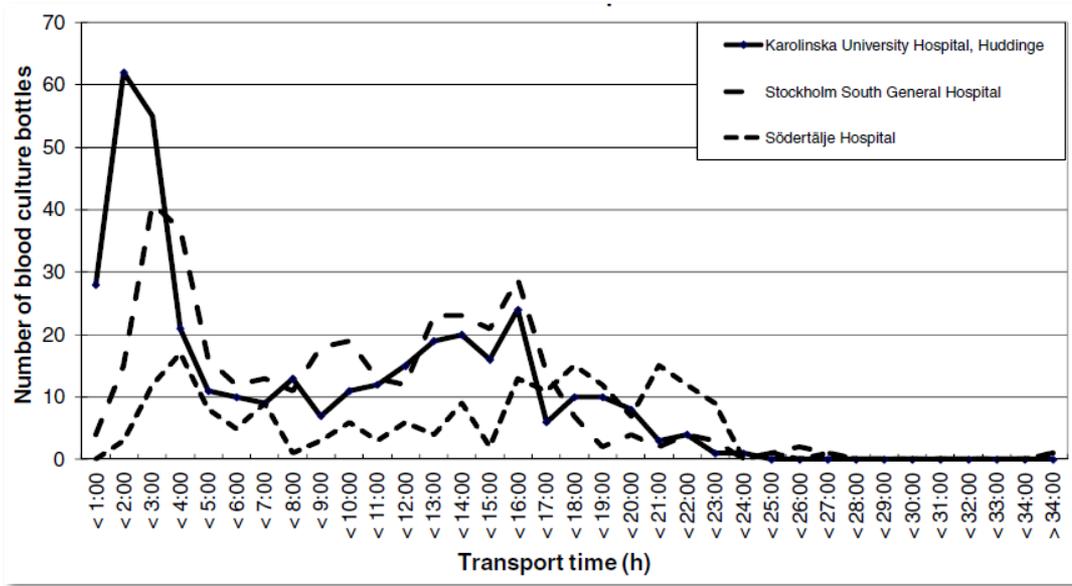
Evgeny A. Idelevich · Gerda Silling · Yvonne Niederbracht · Hanna Penner · Maria Cristina Sauerland · Sascha Tafelski · Irit Nachtigall · Wolfgang E. Berdel · Georg Peters · Karsten Becker · Molecular Diagnostics of Sepsis study group



Sepsis-„Schnell“diagnostik unter Routinebedingungen

(Mo-Fr 7.30 – 18.00)



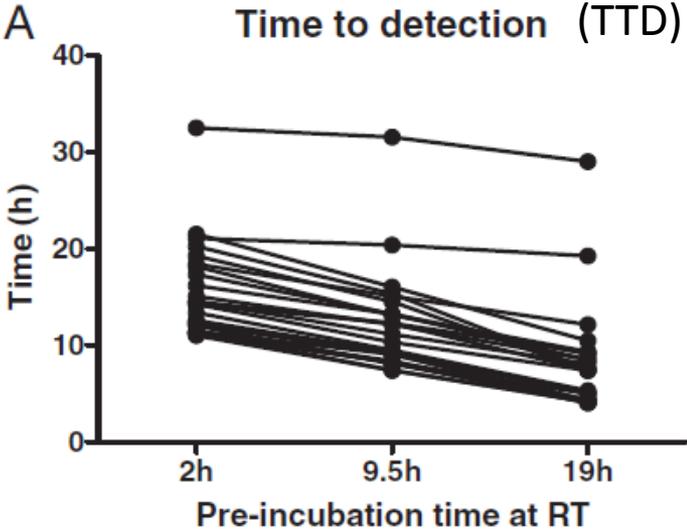


Distribution of transport time for individual blood culture samples

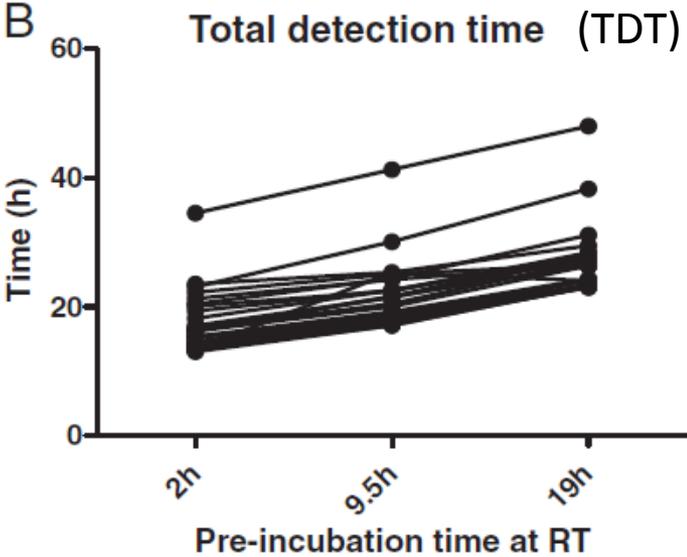
Median (IQR) laboratory transport time for 3 tertiary-care hospitals by time of sampling

	Karolinska University Hospital, Huddinge		Stockholm South General Hospital		Södertälje Hospital	
	No. of samples	Median (IQR)	No. of samples	Median (IQR)	No. of samples	Median (IQR)
Weekdays	274	4 (2–13)	250	8 (3–13)	133	14 (4–18)
Weekends	102	10 (2–15)	98	11 (6–15)	52	17.5 (11–21)
8–16	161	2 (1–3)	117	3 (2–5)	90	18 (4–21)
16–24	154	13 (10–15)	159	13 (11–15)	70	16 (13–17)
24–8	61	6 (4–7)	72	6 (4–8)	25	7 (4–9)

IQR = interquartile range.



Time difference between the time point at which the samples are **placed in the blood culture system** and the time point at which the blood culture bottle **signals for positivity**



Time difference between the **time point of sampling** and the time point at which the blood culture bottle **signals for positivity**. (Transport time + TTD)

- Unfortunately, the shorter TTD obtained with longer transport times **did not compensate** the time period outside the blood culture system
- **TDT** invariably **increased with longer transport time.**

TTD (A) and TDT(B) of single bacterium in simulated blood culture bottle after three different pre-incubation periods.

RT: room temperature

Commercially available systems for the direct identification of microbes from positive BCs

System (Manufacturer)	Methods	Time to result	Microorganism coverage	Resistance and virulence markers	Sensitivity Specificity Correlation with conventional methods (%)
PNA FISH and QuickFISH (AdvanDx, Woburn, MA, USA)	FISH	<1–3 hours	4 Gram positive 4 Gram negative 5 Fungi	0	97–100 90–100 96–99
AccuProbe (Gen-Probe, San Diego, CA, USA)	FISH	<1 hour	<i>Staphylococcus aureus</i> <i>Enterococcus</i> spp. <i>Streptococcus pneumoniae</i> <i>Streptococcus</i> group A <i>Streptococcus</i> group B	0	80.8–100 98.7–100 nr
Verigene (Nanosphere, Northbrook, IL, USA)	Microarray	2.5 hours	12 Gram positive 9 Gram negative	<i>mecA</i> , <i>vanA/B</i> , KPC, NDM, CTX-M, VIM, IMP, OXA12	81–100 98–100 nr
Prove-it Sepsis (Mobidiag, Esbo, Finland)	Microarray	3.5 hours	60 bacteria 13 fungi	<i>mecA</i>	95% 99% nr
FilmArray (Idaho Technology, Salt Lake City, UT, USA)	Multiplex PCR	1 hour	8 Gram positive 11 Gram negative 5 Fungi	<i>mecA</i> , <i>vanA/B</i> , KPC	97–95 91–98 nr
Xpert MRSA/SA BC (Cepheid, Sunnyvale, CA, USA)	Real-time PCR	1 hour	<i>S. aureus</i>	<i>mecA</i>	100 99–100 nr
StaphSR assay (BD GeneOhm, San Diego, CA, USA)	Multiplex PCR	1–2 hours	<i>S. aureus</i>	<i>mecA</i>	96–100 95–98 nr
StaphPlex (Genaco Biomedical Products, Huntsville, AL, USA)	Multiplex PCR + Microarray	5 hours	<i>S. aureus</i>	<i>mecA</i> (+ PVL)	100 95–100 92
MALDI-TOF MS Bruker Daltonics (Bremen, Germany) bioMérieux (Marcy l'Etoile, France)	Mass-spectrometry	<1 hour	<1000 ^a	not in routine	– – 76–99



www.atitesting.com



www.sas.upenn.edu

Semiautomated blood culture instrument (≥1 – 7 days)



Conventional cultivation on solid agar medium (≥1 – 2 days)



www.biomerieux.pl

Biochemical procedures and susceptibility testing (≥1 – 2 days)



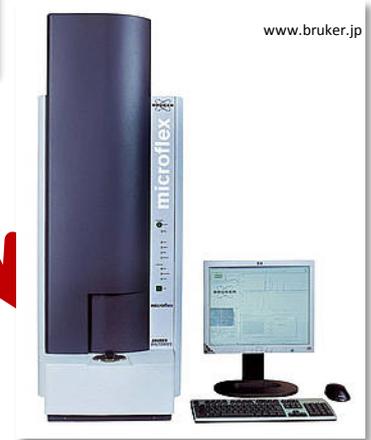
MALDI Sepsityper®

• MALDI Biotyper®

For Direct Pathogen Identification from Positive Blood Culture Bottles

53.1 min
(mean time)

- 6.3 min for lysis-centrifugation
- 32.2 min for extraction and pellet drying
- 11.2 min for spotting
- 3.4 min for MALDI-TOF MS measurement



www.bruker.jp

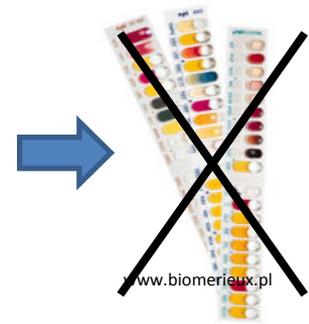
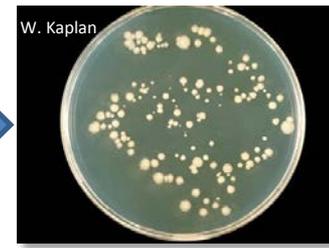
Applied method **saved on average 23.5 h** for yeast identification

Evaluation of Three Rapid Diagnostic Methods for Direct Identification of Microorganisms in Positive Blood Cultures

Raquel M. Martinez,* Elizabeth R. Bauerle, Ferric C. Fang, Susan M. Butler-Wu

Definitive ID	No. of cultures	No. (%) of cultures correctly identified by:			
		MALDI-TOF MS (%)		<i>QuickFISH</i> (%)	BC-GP (%)
		Genus (>1.6)	Species (>1.8)		
Staphylococci	58	57 (98.3)	51 (87.9)	57 (98.3)	58 (100)
Enterococci	16	16 (100)	16 (100)	16 (100)	16 (100)
<i>Enterobacteriaceae</i>	35	34 (97.1)	32 (91.4)	33 (94.3)	—
Non- <i>Enterobacteriaceae</i>	11	6 (54.5)	5 (45.5)	11 (100)	—
Yeast	5	3 (60)	3 (60)	5 (100)	—

- MALDI-TOF MS with Sepsityper (Bruker) processing
- *QuickFISH* (AdvanDx)
- Verigene Gram-Positive Blood Culture (BC-GP; Nanosphere)



Semiautomated blood culture instrument (≥1 – 7 days)

Conventional cultivation on solid agar medium (≥1 – 2 days)

~~Biochemical procedures and susceptibility testing (≥1 – 2 days)~~

- zusätzliche Kosten
- personalaufwendig
- schwierig in die Laborroutine integrierbar

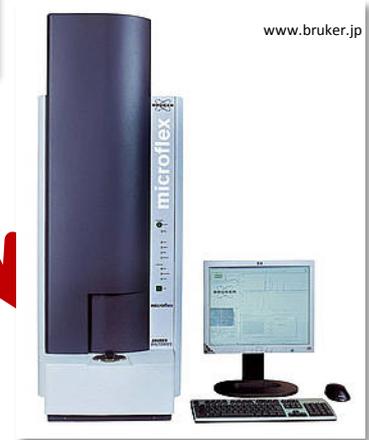


MALDI Sepsityper®

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For Direct Pathogen Identification from Positive Blood Culture Bottles

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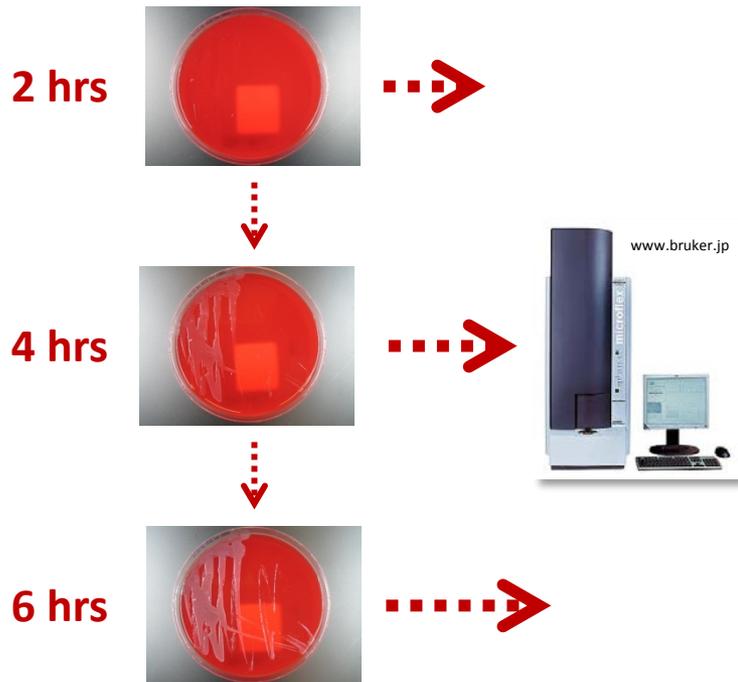
Applied method saved on average 23.5 h for yeast identification

Rapid identification of microorganisms from positive blood cultures by MALDI-TOF mass spectrometry subsequent to very short-term incubation on solid medium

E. A. Idelevich, I. Schüle, B. Grünastel, J. Wüllenweber, G. Peters and K. Becker

Institute of Medical Microbiology, University Hospital Münster, Münster, Germany

Clin Microbiol Infect 2014; **20**: 1001–1006



Cultivation times to successful identification:

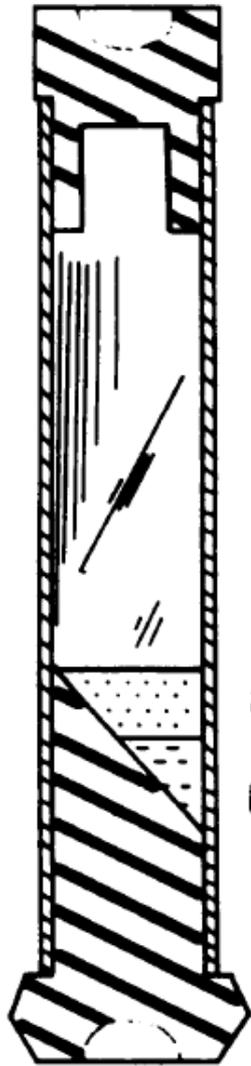
	Species level (score ≥ 2) (%)	Genus level (score ≥ 1.7) (%)
Gram-positive aerobic cocci, $n = 86$		
≤ 2 h	1 (1.2)	5 (5.8)
< 4 h	16 (18.6)	42 (48.9)
< 6 h	55 (64.0)	77 (89.5)
≤ 8 h	83 (96.5)	84 (97.7)
≤ 12 h	85 (98.8)	85 (98.8)
24 h	86 (100)	86 (100)
Mean identification time, h	5.9	4.7
Gram-negative aerobic rods, $n = 42$		
< 2 h	32 (76.2)	37 (88.1)
≤ 4 h	40 (95.2)	41 (97.6)
≤ 6 h	41 (97.6)	41 (97.6)
≤ 8 h	41 (97.6)	41 (97.6)
≤ 12 h	41 (97.6)	41 (97.6)
24 h	41 (97.6)	41 (97.6)
Mean identification time, h	2.0	1.7

- Very early and reliable identification from positive BCs
- Without additional time and cost expenditure

New Centrifugation Blood Culture Device

G. L. DORN* AND K. SMITH

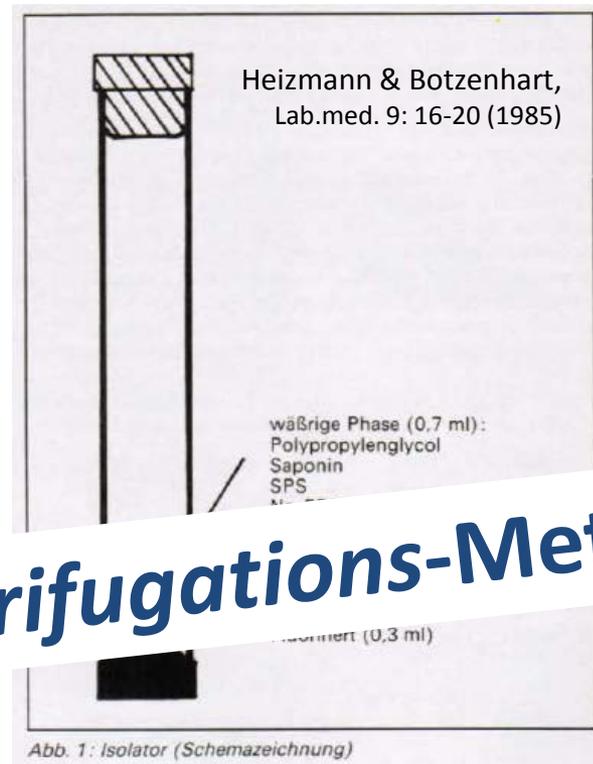
Department of Microbiology, Leland Fikes Research Institute, and Granville C. Morton Cancer and Research Hospital, Divisions of Wadley Institutes of Molecular Medicine, Dallas, Texas 75235



Vacuum chamber

Hemolyzing and
anticoagulating agent

Cushion



Lysis-Zentrifugations-Methode

and time. The best overall average recovery (93%) was obtained at $3,000 \times g$ for 30 min. With

⇒ **qualitatives und quantitatives Blutkulturverfahren**

Direct blood culturing on solid medium outperforms an automated continuously monitored broth-based blood culture system in terms of time to identification and susceptibility testing

E. A. Idelevich, B. Grünastel, G. Peters and K. Becker

New Microbe and New Infect 2016; 10: 19–24



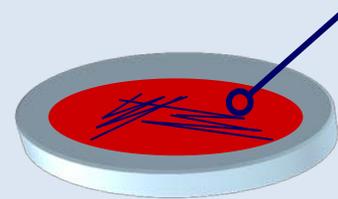
10 ml of spiked blood



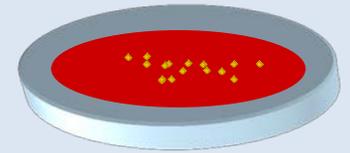
Isolator tube



Centrifugation (3000 g for 30 min)

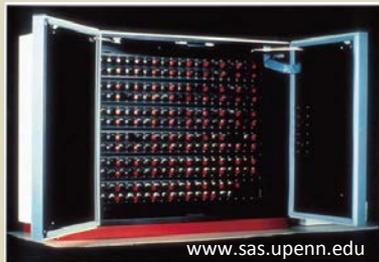


Incubation at 36°C in air with 5% CO₂



(neu)

Hourly observation of growth for **microcolonies**



Automated BC system (BACTEC™ 9240; BD Diagnostics)

MALDI-TOF MS (Microflex LT system; Bruker Daltonics)



www.bruker.jp

Direct blood culturing on solid medium outperforms an automated continuously monitored broth-based blood culture system in terms of time to identification and susceptibility testing

E. A. Idelevich, B. Grünastel, G. Peters and K. Becker

New Microbe and New Infect 2016; 10: 19–24

Organism	Concentration (CFU/mL)	Time to MALDI-TOF MS identification after direct blood culture method (h)			Time to BACTEC positivity sign h
		High confidence (score ≥ 2.0)	Low confidence (score ≥ 1.7)	Modified threshold (score ≥ 1.5 and three identical propositions)	
<i>Escherichia coli</i>	100	7	7		
	10	7			
<i>Pseudomonas aeruginosa</i>	100	7	7		14.0
<i>Staphylococcus aureus</i>	100	7	7		13.9
<i>Staphylococcus pneumoniae</i>	100	7	7		16.1
<i>Enterococcus faecalis</i>	100	15	15	14	17.8
<i>Enterococcus faecium</i>	100	9 (8 ^a)	8	8	10.7
<i>Streptococcus pneumoniae</i>	10	9	9	9	11.9
<i>Streptococcus pneumoniae</i>	1	10	10	10	13.0
<i>Streptococcus pneumoniae</i>	100	9	8	8	10.2
<i>Streptococcus pneumoniae</i>	10	9	9	9	11.5
<i>Streptococcus pneumoniae</i>	1	12	12	12	12.4
<i>Haemophilus influenzae</i>	100	11	11	11	16.3
	10	11	11	11	18.6
	1	11	11	11	20.0

⇒ Spezies-Identifikation durchschnittlich 3,3 h eher als Wachstumsdetektion durch automatisiertes System

^aTime to identification was reduced by on-plate extraction.

^bTime to identification was reduced by on-plate extraction followed by additional shots.

^cNo Vitek 2 antimicrobial susceptibility testing is routinely performed for *H. influenzae*.

Direct blood culturing on solid medium outperforms an automated continuously monitored broth-based blood culture system in terms of time to identification and susceptibility testing

E. A. Idelevich, B. Grünastel, G. Peters and K. Becker

New Microbe and New Infect 2016; 10: 19–24

Organism	Concentration (CFU/mL)	Time to Vitek 2 AST after direct blood culture method (h)			Time to BACTEC positivity signal, h
		Time until Vitek 2 inoculation	Duration of Vitek 2 AST	Total time to Vitek 2 result	
<i>Escherichia coli</i>	100	7			
	10				
<i>Staphylococcus aureus</i>	100	14	11.8	19.8	12.7
	10	14	11.8	25.8	14.0
	1	17	13	30.0	16.1
<i>Enterococcus faecalis</i>	100	8	11.5	19.5	10.7
	10	9	11.8	20.8	11.9
	1	10	12	22.0	13.0
<i>Streptococcus pneumoniae</i>	100	9	13.8	22.8	10.2
	10	10	13.8	23.8	11.5
	1	12	10.3	22.3	12.4
<i>Haemophilus influenzae</i>	100	— ^c	—	—	16.3
	10	—	—	—	18.6
	1	—	—	—	20.0

⇒ Empfindlichkeitstestung durchschnittlich 2.5 h eher gestartet als Wachstumsdetektion durch automatisiertes System

^cNo Vitek 2 antimicrobial susceptibility testing is routinely performed for *H. influenzae*.

Direkte Blutkultivierung von Hefen auf Festmedium

Organism and concentration	Time to species identification applying direct lysis-centrifugation-cultivation method		Time to positivity applying automated BC system	
	Sabouraud agar	Chocolate agar	BACTEC Mycosis-IC/F	BACTEC Plus Aerobic/F
<i>Candida albicans</i>				
100 cfu/ml	14	15	16.4	17.8
10 cfu/ml	15	18	19.1	21.3
1 cfu/ml	17	20	23.1	24.6
<i>Candida glabrata</i>				
100 cfu/ml	15	26	16.2	38.2
10 cfu/ml	15	27	19.5	47.7
1 cfu/ml	16	30	23.6	56.6
<i>Candida krusei</i>				
100 cfu/ml	15	15	16.9	17.5
10 cfu/ml	15	18	19.3	20.2
1 cfu/ml	17	19	23.0	22.1
<i>Candida tropicalis</i>				
100 cfu/ml	15	13	15.6	16.5
10 cfu/ml	15	16	17.2	19.2
1 cfu/ml	16	17	20.5	20.9

Idelevich *et al*, ECCMID 2015

Rapid Identification and Susceptibility Testing of *Candida* spp. from Positive Blood Cultures by Combination of Direct MALDI-TOF Mass Spectrometry and Direct Inoculation of Vitek 2

Evgeny A. Idelevich*, Camilla M. Grunewald, Jörg Wüllenweber, Karsten Becker

PLOS ONE | DOI:10.1371/journal.pone.0114834 December 9, 2014

Positive monomicrobial BCs from 15 patients including:

- 13 aerobic
- 5 anaerobic
- 5 fungal
- 1 paediatric bottle

Candida species identification rates by using direct MALDI-TOF MS method

Species	Total number of samples	Number of identified samples	Identification rate, %
<i>C. glabrata</i>	12	7	58.3
<i>C. albicans</i>	9	6	66.7
<i>C. dubliniensis</i>	2	1	— ^a
<i>C. krusei</i>	1	1	— ^a
Total	24	15	62.5

Commercially available molecular assays for the diagnosis of sepsis using whole blood as sample type

Diagnostic technique		No. of pathogens detected	Sensitivity	Specificity
Xpert MRSA/SA (Roche)	Real-time PCR	2	75–100	98.4–99.4
SeptiFast (Roche)	Multiplex real time PCR assay for bacterial and fungal pathogens	25, plus mecA as reflex test	60–95	74–99
VYOO (SIRS-Lab)	Multiplex PCR with gel electrophoresis	34plus mec A, vanA/B/C, SHV, CTX-M	30–51	n.d.
SepsiTest (Molzym)	Broad-range PCR with sequencing	>300 pathogens	61–88.5	83.5–85.8

*Concordance with blood culture-dependent assays; n.d., not determined

Liesenfeld et al., Eur. J. Microbiol. Immunol.

Patient outcome and hospital costs for patients with bloodstream infection treated using routine medical management with or without the SeptiFast test

	Routine management (Mean \pm SD)	Routine management plus SeptiFast (Mean \pm SD)	<i>p</i> value
28-day mortality	13 (27%)	14 (26%)	n.s.
6-month mortality	20 (37%)	20 (41.6%)	n.s.
Stay in ICU	31.0 \pm 19.4	22.9 \pm 29.9	<0.05
Stay in hospital	21.3 \pm 23.4	18.3 \pm 21.4	<0.05
Stay in ICU survivors	24.1 \pm 21.9	18.3 \pm 11.4	<0.05
Number of antibiotics used per patient	5.1 \pm 3.1	4.2 \pm 2.2	<0.05
Antibiotic treatment cost per patient	3576 €	2812 €	<0.05
Cost of ICU stay	32798 €	24246 €	<0.05
Cost of ward stay	5824 €	4988 €	<0.05
Total cost	42198 €	32228 €	<0.05

n.s., not significant

acc. data from Alvarez et al., Anaesth. Intensive Care 40, 958–963 (2012)

Liesenfeld et al., Eur. J. Microbiol. Immunol.

Randomized controlled clinical trial evaluating multiplex polymerase chain reaction for pathogen identification and therapy adaptation in critical care patients with pulmonary or abdominal sepsis

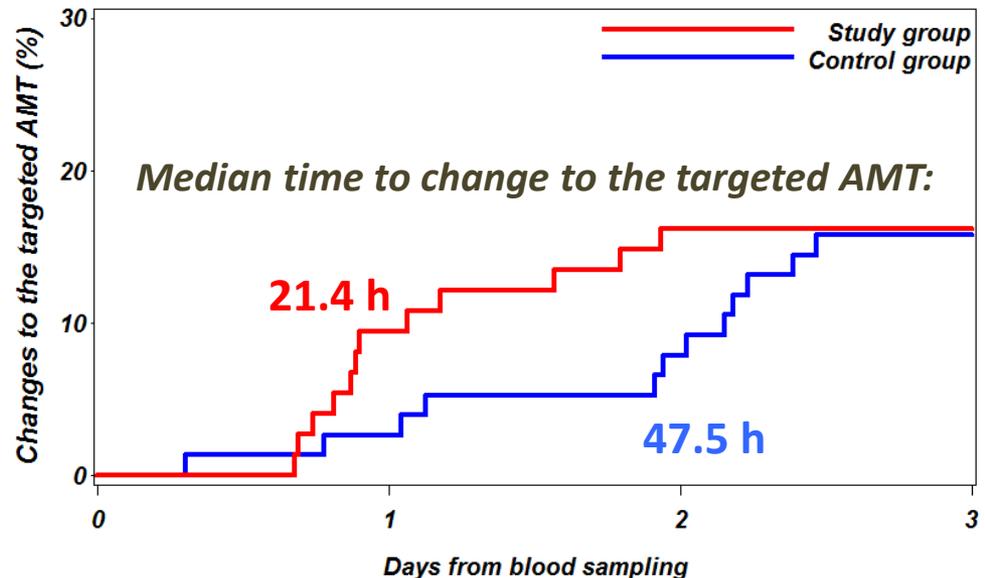
Sascha Tafelski^{1,*}, Irit Nachtigall^{1,*}, Thomas Adam², Stefan Bereswill², Jana Faust¹, Andrey Tamarkin¹, Tanja Trefzer¹, Maria Deja¹, Evgeny A Idelevich³, Klaus-Dieter Wernecke^{4,5}, Karsten Becker³, Claudia Spies¹ for the Molecular Diagnostics of Sepsis Study Group

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ORIGINAL INVESTIGATION

Impact of multiplex PCR on antimicrobial treatment in febrile neutropenia: a randomized controlled study

Evgeny A. Idelevich · Gerda Silling · Yvonne Niederbracht · Hanna Penner · Maria Cristina Sauerland · Sascha Tafelski · Irit Nachtigall · Wolfgang E. Berdel · Georg Peters · Karsten Becker · Molecular Diagnostics of Sepsis study group



- Significant reduction in the time required for initial pathogen identification by PCR (15.9 h), compared with standard BC (38.1 h); ($P < 0.001$) [Tafelski et al.].
- PCR method accelerates change to the targeted antimicrobial therapy (AMT) in febrile neutropenic patients ($P = 0.018$) [Idelevich et al.]

„Wat wellste maache?“

- Definition der Sepsis „in progress“
- ungebrochene, Patientencharakteristika-bedingte Bedeutung der Sepsis bzgl. Morbidität, Letalität und Gesundheitsökonomie
- therapeutische Limitierungen durch (multi-) resistente Erreger
- BK-Diagnostik ist essentiell trotz bestehender Limitationen!
- nur teilweiser Fortschritt bei der Erfüllung der klinischen Anforderungen an die mikrobiologische Diagnostik (sensitiv, spezifisch und **schnell!!!**)
 - inakzeptable Transport- und Lagerungszeiten (Station, Labor)
 - nur partielle Vorteile durch DNA-basiertem Direktnachweis – bisher kein Durchbruch in der Sepsis-Routinediagnostik
 - zu lange Kultivierungszeiten
 - erfolgreiche Beschleunigung der Zeiten für Identifizierung und Empfindlichkeitstestung durch
 - MALDI TOF MS
 - Einsatz von Mikrokolonien nur kurzzeitig bebrüteter Festnährmedien

„Wat ~~wollste~~ maache?“
sollste

⇒ **Beschleunigung**
der mikrobiologischen
Sepsisdiagnostik